

# Fast-field cycling NMR is sensitive to the method of cross-linking in BSA gels

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## Introduction

In contrast to conventional nuclear magnetic resonance (NMR) experiments where a static magnetic field is applied to a sample, the applied field in field-cycled NMR is altered during the experiment. Field-cycling allows measurements, for example of the spin-lattice relaxation rate ( $R_1$ ), to be made as a function of the applied field. At several discrete field strengths the  $^1\text{H}$  NMR and  $^{14}\text{N}$  NQR frequencies coincide, allowing effective relaxation of the magnetisation from bulk water protons *via* the quadrupolar  $^{14}\text{N}$  nucleus [1]. Therefore, in a plot of  $R_1$  versus field, ‘quadrupolar peaks’ are often observed at these field strengths.

It has been suggested that quadrupolar peaks in proteinous samples result from interactions between sufficiently-bound nitrogenous functional groups and low-mobility water protons [2]. Furthermore, the amplitude of the quadrupolar peak has been shown to be proportional to protein concentration [3]. In this study, the quadrupolar peaks of gels of bovine serum albumin (BSA) formed by boiling or chemical cross-linking were examined.

## Methods

BSA gels (final concentration 9% w/v) in 20.5 mM bicarbonate buffer (pH 7.3) were prepared by boiling or addition of formalin or glutaraldehyde (final concentration 12.5% w/v). Measurements of  $R_1$  were made between 0.047–187.89 mT at 37 °C on a SMARtracer relaxometer (Stelar S.r.l., Mede, Italy). A power-law with Lorentzian-bell algorithm, derived from the literature [3,4], was fit to the data (Matlab 2012a, The Mathworks, Cambridge, UK; scripts developed by Lionel M. Broche, Aberdeen, UK).

## Results

Dispersion curves were dependent on the method of gel formation (Figure 1). The quadrupolar peak amplitude was largest with formaldehyde (Figure 2A). The eta-value, describing the shape of the quadrupolar peaks, was similar with chemical cross-linking but larger if boiled (Figure 2B).

## Discussion

Boiled BSA gels likely contain denatured protein in a network of fragments, monomers and higher aggregates [5] linked by various functional groups (e.g. amino, amide and sulphhydryl groups). Glutaraldehyde gels may contain BSA aggregates in a native configuration *via* the majority of free amino-groups [6]. Formaldehyde gels are likely cross-linked in a heterogeneous manner, involving fewer free amino-groups [7,8]. Therefore, quadrupolar peak amplitude and shape may be related to the macroscopic protein network, and/or the functional groups involved in gel cross-linking. These observations may be pertinent in the analysis of fixed tissue samples by field-cycled NMR, where any observed quadrupolar signals may be affected by the method of tissue fixation.

## Acknowledgements

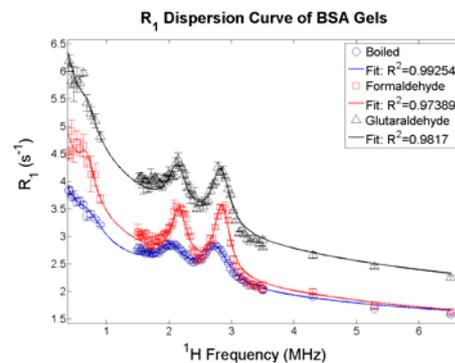
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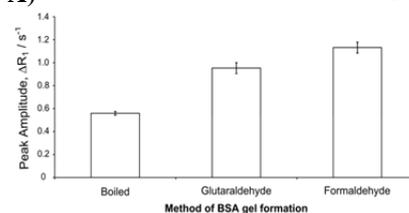
## References

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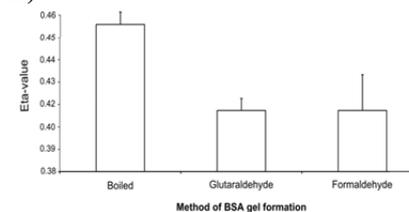


**Figure 1:** Dispersion curves of BSA gels generated by different cross-linking methods.

### A) Amplitude of the observed quadrupolar peak in the BSA gel



### B) Eta-value of the observed quadrupolar peak in the BSA gel



**Figure 2:** A) Amplitude and B) eta-value of the quadrupolar peak of cross-linked BSA gels.