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RESEARCH PAPER

Association of heat shock protein70-2 (*HSP70-2*) gene polymorphism with obesity

Maryam Mardan-Nik^{1*}, Alireza Pasdar^{2,3*}, Khadijeh Jamialahmadi^{4,5*}, Amir Avan², Mohsen Mohebbati¹, Habibollah Esmaily⁶, Atefeh Biabangard-Zak⁷, Fahimeh Afzal Javan⁸, Mahdi Rivandi⁸, Gordon A. Ferns⁹, and Majid Ghayour-Mobarhan¹⁰

¹Cardiovascular Research Center, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran, ²Department of Modern Sciences and Technologies, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran, ³Division of Applied Medicine, Medical School, University of Aberdeen, Foresterhill, Aberdeen, UK, ⁴Department of Medical Biotechnology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran, ⁵Biotechnology Research Center, Mashhad University of Medical Sciences, Mashhad, Iran, ⁶Department of Biostatistics and Epidemiology, School of Health, Mashhad University of Medical Sciences, Mashhad, Iran, ⁷Department of Biochemistry, Golestan University, Gorgan, Iran, ⁸Student Research Committee, Department of Modern Sciences & Technologies, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran, ⁹Brighton & Sussex Medical School, Division of Medical Education, Falmer, Brighton, Sussex, UK, ¹⁰Biochemistry of Nutrition Research Center, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

Abstract

Background: Obesity is a major risk factor of chronic-diseases, including cardiovascular-diseases (CVD). Increasing evidence is showing the association of heat-shock protein (HSP) with type-2 diabetes and CVD; however, there is little data on the relationship between the genetic-polymorphisms of *HSP70-2* with obesity.

Aim: The present study has investigated the association between *1267HSP70-2* genetic polymorphism and obesity in an Iranian population with 317 subjects.

Subjects and methods: Anthropometric parameters and biochemical measurements were measured in all the samples, while genotypes were determined using PCR-RFLP. Univariate/multivariate analyses were conducted to explore the relationship between the genetic-polymorphisms and obesity.

Results: The data showed a significant association between *1267HSP70-2* polymorphism in obese subjects, compared to the non-obese group. Moreover, it was observed that this polymorphism was associated with obesity in the CAD+group, which had a high BMI compared to non-obese controls.

Conclusion: The *1267HSP70-2* polymorphism is associated with obesity in an Iranian population, supporting a possible potential genetic link between obesity and cardiovascular diseases.

Keywords

Obesity, heat shock protein 70, PCR-RFLP, polymorphism, coronary artery disease

History

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Introduction

Obesity is one of the major risk factors of metabolic and chronic diseases and its prevalence is increasing worldwide (Jou & Techakehajib, 2012; Sakowicz et al., 2013). Several studies have identified a number of genetic variants associated with obesity (Yu et al., 2012). In the past, genetic studies of obesity were concentrated on the rare monogenic syndromes. However, the associations between genetic polymorphisms

and obesity are frequently reported as heterogeneous (Bienertova-Vasku et al., 2008; Paracchini et al., 2005).

Heat shock proteins (HSPs) belong to a multi-gene family and their molecular weight varies from 8–150 kDa (Whitley et al., 1999). Heat and several other stimuli such as cold, UV radiation, alcohol, heavy metal ions, oxidation, low glucose, infection, inflammation and hypoxia can affect heat shock protein expression (Kiang & Tsokos, 1998). Heat shock proteins are typically named and classified according to their molecular size. *HSP70*, a 70 kDa protein, is coded by *HSP70* gene (Whitley et al., 1999). Three genes that encode members of the *HSP70* class are *HSP70-1* (*HSPA1*), *HSP70-2* (*HSPA1B*) and *HSP70-hom* (*HSPAIL*). All three of these genes are located in the region of MHC class III (Sargent et al., 1989). *HSP70-1* and *HSP70-2* code similar proteins with 641 amino acids (Milner & Campbell, 1990).

Some studies have shown that inflammation and high levels of serum lipids could result in increased accumulation of lipids in the artery wall. Accumulation of lipids induces the secretion

*These authors contributed equally to the study.

Correspondence: Majid Ghayour-Mobarhan, MD, PhD, Biochemistry of Nutrition Research Center, Faculty of Medicine, Mashhad University of Medical Science, Mashhad, Iran, 99199-91766. Tel: +985118002288. Fax: +985118002287. E-mail: ghayourm@mums.ac.ir

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of inflammatory cytokines such as tumour necrosis factor- α (TNF- α) from macrophages and adipocytes. TNF plays a role in activation of the serine threonine kinase, namely JNK (c-jun amino terminal kinase) and IKK (inhibitor of B kinase) in insulin-responsive (Watt et al., 2006). JNK and IKK can lead to insulin resistance by phosphorylation of the IRS-1 (Insulin Receptor Substrate 1) (Hotamisligil, 2006) and *HSP70-2* expression can inhibit the activation of the JNK and IKK. Induction of *HSP70-2* prevents insulin resistance and, thus, is regarded as a potential target in the prevention and treatment of obesity and type 2 diabetes (Chung et al., 2008; Hooper & Hooper, 2009). These potential properties suggest the *HSP70* gene may be a candidate in the aetiology of obesity.

There is a growing body of evidence showing that this gene is associated with type 2 diabetes (Zouari Bouassida et al., 2004) and coronary artery disease (Hrira et al., 2012). Moreover, in our recent study we reported an association between the *HSP70-2* gene +1267A>G polymorphism and cardiovascular disease in 628 Iranian patients (Mardan-Nik et al., 2014). In the present study we have investigated further the relationship between *HSP70-2* gene polymorphisms and obesity in an Iranian population with 317 subjects.

Materials and methods

Study population

The study population consisted of 317 patients (57.4% male and 42.6% female, aged 35–78 years) who underwent coronary angiography in the Ghaem Hospital Medical Center, Mashhad, Iran (Zomorrodian et al., 2015). Of the total group of patients, 94 were obese (BMI \geq 30). Control subjects ($n = 233$, 86.1% male and 13.9% female, aged 37–67 years) were healthy volunteers (93 obese and 130 non-obese). Informed consent was taken from all the participants. The study was approved by the Ethics Committee of the Mashhad University of Medical Sciences (MUMS).

Biometric and biochemical measurements

Anthropometric parameters of individuals including weight, height, BMI, waist circumference, hip circumference and waist/hip ratio as well as systolic and diastolic blood pressures were measured as previously described (Emamian et al., 2015; Ghayour-Mobarhan et al., 2008; Mirhafez et al., 2015a). Obesity was defined according to the World Health Organisation (WHO) (Farshidi et al., 2010).

Biochemical analysis

A full fasted lipid profile was determined for each subject. Serum lipids and fasting blood glucose (FBG) concentrations were measured by enzymatic methods as previously described (Ghayour-Mobarhan et al. 2008; Mirhafez et al. 2015b; Oladi et al., 2015).

DNA isolation and SNP selection and genotyping

Genomic DNA was extracted from peripheral blood using a commercially available DNA isolation kit (Genet bio, Daejeon, Korea) according to the manufacturer's protocol. The quality of the DNA (ng/ μ l) samples was assessed using agarose gel-electrophoresis and the concentration quantities

by spectrophotometry (Nano Drop 1000, Thermo Scientific, Wilmington, NC). Genotyping for the rs1061581 polymorphism was carried out using polymerase chain reaction–restriction fragment length polymorphism (PCR-RFLP) technique. Amplification of *HSP70-2* gene was performed using forward and reverse primers as follows: sense 5'-CATCGACTTCTACACGTCC-3' and antisense 5'-CGGAGTAGGTGGTGAAGATC-3'. The PCR reaction was performed in 25 ml final volume, using 100 ng of genomic DNA, 0.2 mM dNTPs, 2 mM MgCl₂, 1 \times Taq DNA polymerase buffer, 0.32 pmol each primer and 1 unit of Taq DNA polymerase (Genet bio, Korea). The polymerase chain reaction conditions were as follows: initial denaturation at 95°C for 5 minutes, followed by 30 cycles denaturation at 95°C for 30 seconds, annealing at 61°C for 30 seconds, DNA extension at 72°C for 1 minute and the final extension at 72°C for 7 minutes. All amplification cycles were performed in PCR system Verity 96 well thermocycler (Applied Biosystems, CA). PCR products were digested for 16 hours at 37°C with 1 μ l of *Pst*I (Fermentas, Vilnius, Lithuania). For separation of RFLP products, electrophoresis on 1.5% agarose gel with ethidium bromide staining and visualising by UV light was performed. DNA lacking polymorphic *Pst*I site (adenine at 1267 nt) within the *HSP70-2* gene produced a fragment of 428 bp (A allele), whereas the presence of 1267 G allele generated two fragments of 247 and 181 bp after *Pst*I digestion (G allele). Finally, a direct sequencing approach (Company of Sequetech, CA) was used to confirm the genotypes obtained by PCR-RFLP for some of the subjects in different groups (Figure 1).

Statistical analyses

All data were analysed using the SPSS for Windows, version 22 software package (SPSS Inc, Chicago, IL). The Kolmogorov–Smirnov test was used to test the normality of

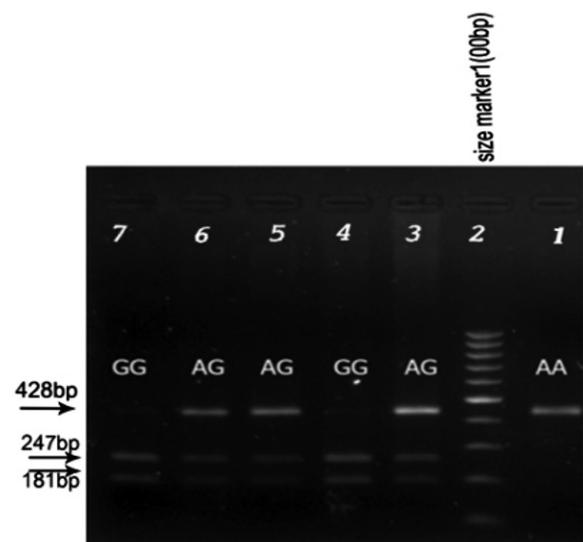


Figure 1. PCR-RFLP of the amplified segment in 1267*HSP70-2* gene. The genotype was labelled on corresponding sequences and the sites which were marked with black arrows were the SNP of *HSP70-2* gene. Electrophoresed on 1.5% agarose, stained with ethidium bromide. Lane 2, 100bp DNA ladder. Lane 3, 5 and 6 heterozygote for *HSP70-2* genotype, Lane 4 and 7 homozygote GG for *HSP70-2* genotype. Lane 1 homozygote AA for *HSP70-2* genotype.

Table 1. Demographic and clinical characteristics of the subjects.

Characteristics	Non-obese control (n = 130)	Obese control (n = 93)	Obese case (n = 94)	Comparison between the groups		
				P1	P2	P3
Age, year	53 (12)	52 (14)	62 (21)	—	*	*
Gender, n (%) Male	110 (84.6)	82 (88.2)	54 (57.4)	—	*	*
BMI (Kg/m ²)	26.3 (4.7)	32.3 (3.50)	32 (4.86)	*	*	—
Weight (Kg)	67 (13)	80 (17.5)	80 (15)	*	*	—
WC (cm)	94 (12)	107 (13)	103 (18)	*	*	—
Height (cm)	161 (12)	157 (15)	81 (14)	*	*	—
TC (mg/dl)	188 (73)	193 (86)	184 (98)	—	*	*
TG (mg/dl)	116 (83)	140 (85)	141 (140)	—	*	—
HDL (mg/dl)	42 (10.9)	42 (12)	38 (12.5)	—	*	*
LDL (mg/dl)	120 (45)	113 (38)	116 (70)	—	*	*
FBG (mg/dl)	80 (17)	84 (26)	96 (91)	—	*	*
HC (cm)	100 (7)	110 (10)	105 (23)	*	*	*
Waist/Hip ratio	0.94 (0.09)	0.95 (0.09)	0.99 (0.15)	—	*	*
SBP (mmHg)	123 (28)	123 (28)	150 (30)	—	*	*
DBP (mmHg)	80 (8)	80 (10)	80 (15)	—	—	—
hs-CRP	2.42 (2.4)	1.99 (3.06)	2.82 (6.75)	—	*	*
Hypertension, n (%)	211 (16.3)	31 (33.7)	61 (64.9)	*	*	*
Diabetes, n (%)	10 (7.9)	10 (11)	40 (42.6)	—	*	*

Values are expressed as interquartile range non-normally distributed variables. Comparisons were performed by independent samples *t*-test and Mann–Whitney U-test for non-normally distributed variables.

χ^2 of test results for categorical data.

BMI, body mass index; WC, waist circumference; TC, total cholesterol; TG, triglycerides; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; FBG, fasting blood glucose; HC, hip circumference; SBP, systolic blood pressure; DBP, diastolic blood pressure; P1, comparison between groups of non-obese control and obese control; P2, comparison between groups of non-obese control and obese case; P3, comparison between groups of obese control and obese case.

Table 2. Genotype distribution and allele frequencies of 1267HSP70-2 polymorphism in non-obese and obese control groups.

	Non-obese control (n = 130)	Obese control (n = 93)	OR (95% CI)	<i>p</i> -value	Adj. OR (95% CI)	Adj. ^a <i>p</i> -value
AA, n (%)	46 (35.4)	17 (18.3)	0.304 (0.124–0.749)	0.010	0.346 (0.120–3.995)	0.049
AG, n (%)	70 (53.8)	59 (63.4)	0.694 (0.316–1.526)	0.364	0.898 (0.332–2.429)	0.832
GG, n (%)	14 (10.8)	17 (18.3)	1			
A allele	162 (62.3)	93 (50.0)	0.605 (0.413–0.886)	0.01	0.589 (0.366–0.949)	0.030
G allele	98 (37.3)	93 (50.0)	1			

CI, confidence interval; OR, odds ratio; TG, triglycerides.

^aAdjusted for weight.

the variables in each group. Data were expressed as median and interquartile range (IQ₃–IQ₁) for data with non-normal distribution. χ^2 test was used for categorical data. The statistical difference in genotype distribution and allele frequencies between groups was assessed by the χ^2 test. Compliance of genotypes with the Hardy-Weinberg equilibrium in each group was also assessed by χ^2 test. Binary logistic regression was used to adjust for confounders. A 2-sided *p* < 0.05 was considered significant.

Results

Demographics and clinical characteristics of population

The demographic and metabolic characteristics of study subjects with and without obesity are presented in Table 1. The proportion of individuals who had coronary artery disease and obesity was 57.4% and 42.6% in males and females, respectively. The frequency of obesity observed in male patients was significantly higher compared to the female individuals (*p* < 0.001). Total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol

(HDL-C), hip circumference, waist/hip ratio, hs-CRP and FBS were significantly different between patients and control subjects with obesity. There was also no significant difference in the weight, waist circumference, BMI and TG: triglyceride concentrations (*p* > 0.05). The prevalence of diabetes mellitus and hypertension was significantly higher in the obese case group compared to the obese control group (Table 1).

There was a statistically significant difference for anthropometric and biochemical parameters between obese cases and non-obese control subjects except DBP. As would be expected, there were statistically significant differences for hypertension and diabetes between non-obese controls and obese cases (Table 1).

Association between 1267HSP70-2 polymorphism and obesity

The distribution of 1267Hsp70-2 genotypes and the allelic frequencies in study subjects are shown in Tables 2–5. There was a deviation from the Hardy-Weinberg equilibrium in both obese cases and controls with obesity (*p* < 0.05). To rule out any genotyping errors, 10% of samples were genotyped. The frequencies of AA, AG and GG genotypes were 35.4%, 53.8%

Table 3. Dominant analysis model of HSP70-2 gene + 1267A > G polymorphism in non-obese control and obese control groups.

	Non-obese control (n = 130)	Obese control (n = 93)	p-value	OR (95% CI)	Adj. ^a p-value	Adj. OR (95% CI)
AA, n (%)	46 (35.4)	17 (18.3)	1			
AG-GG, n (%)	84 (64.1)	76 (81.7)	0.006	2.448 (1.295–4.629)	0.009	2.790 (1.288–6.047)

CI, confidence interval; OR, odds ratio.

^aAdjusted for weight; TG, triglycerides.

Table 4. Genotype distribution and allele frequencies of 1267HSP70-2 polymorphism in non-obese control and obese case group.

	Non-obese control (n = 130)	Obese case (n = 94)	OR (95% CI)	p-value	Adj. OR (95% CI)	Adj. ^a p-value
AA, n (%)	46 (35.4)	14 (14.9)	0.266 (0.105–0.678)	0.005	0.107 (0.021–0.546)	0.007
AG, n (%)	70 (53.8)	64 (68.1)	0.800 (0.362–1.769)	0.581	0.731 (0.244–2.192)	0.576
GG, n (%)	14 (10.8)	16 (17)	1			
A allele	162 (62.3)	92 (48.9)	0.580 (0.396–0.848)	0.005	0.478 (0.278–0.820)	0.007
G allele	98 (37.3)	96 (51.1)	1			

CI, confidence interval; OR, odds ratio.

^aAdjusted for age ≥ 60; sex; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; FBG, fasting blood glucose; Waist/Hip ratio.

Table 5. Dominant analysis model of HSP70-2 gene + 1267A > G polymorphism non-obese control and obese case group.

	Non-obese control (n = 130)	Obese case (n = 94)	p-value	OR (95% CI)	Adj. ^a p-value	Adj. OR (95% CI)
AA, n (%)	46 (35.4)	14 (14.9)	1			
AG-GG, n (%)	84 (64.1)	80 (85.1)	0.001	3.129 (1.598–6.128)	0.002	4.898 (1.763–13.604)

CI, confidence interval; OR, odds ratio.

^aAdjusted for age ≥ 60; sex; HDL-C; high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; FBG, fasting blood glucose; Waist/Hip ratio.

and 10.8% in the non-obese control and 14.9, 68.1% and 17% in obese cases, respectively. There was a significant association in genotype distribution between the two groups ($p = 0.002$). Compared with the GG genotype of HSP70-2 gene, subjects carrying the AA genotype showed a decreased risk in non-obese controls ($p = 0.005$, OR = 0.26, 95% CI = 0.105–0.678). Similar values were obtained after adjusting for individuals older than 60 years, sex, LDL-C, FBG, HDL and waist/hip ratio. There was a significant difference in frequency of the A allele between two groups ($p = 0.005$, OR = 0.580, 95% CI = 0.396–0.848) (Table 4). The frequencies of the AA, AG and GG genotypes were also 14.9%, 68.1% and 18.3% in the obese cases and 18.3%, 63.4% and 18.3% in the obese controls, respectively. In a dominant analysis model of the HSP70-2 gene + 1267A/G position (AA vs AG + GG), the percentage of subjects who were either homo- or heterozygous for the G allele (1267AG and 1267GG) was significantly higher in obese cases than non-obese controls (85.1% vs 64.1%) (adj. $p = 0.002$, adj. OR = 4.898; 95% CI = 1.763–13.604) (Table 5).

Association between HSP70-2 rs1061583 polymorphism and clinical-biochemical parameters

We also examined the baseline characteristics between the genotype groups in overall groups, but there was no significant difference except SBP and DBP in non-obese controls (data not shown).

Discussion

The complex aetiology of obesity reflects effects of genes and environment as well as their interactions (Bienertova-Vasku et al., 2008). The HSP70-2 gene encodes a protein involved in the pathophysiology of obesity and diabetes (Bouchard, 2008; Chouchane et al. 2001; Chung et al., 2008). In this study it was hypothesised that it might be associated with the prevalence of obesity in Iranian population patients with CAD.

To the best of our knowledge this is the first study evaluating the association of 1267HSP70-2 polymorphism with the prevalence of obesity in Iranian patients with CAD. Our results showed a significant decrease of AA genotype in the obese case group when compared with the G allele carriers (GG and AG) ($p = 0.005$, OR = 0.26, 95% CI = 0.105–0.678), suggesting that this genotype may be considered as a protective marker in the non-obese control group. This observation is consistent with previous results that indicated GG genotype in Tunisians was correlated with obesity (Chung et al., 2008). A recent report by Zouari Bouassida et al. (2004) demonstrated that the 1267 HSP70-2G > A variant could convey an increased risk for obesity and type 2 diabetes. In addition to the single locus analysis our study indicated that there was no association between 1267HSP70-2 polymorphism in both cases and controls with obesity ($p = 0.77$). However, allelic and genotypic frequencies for the polymorphism of HSP70-2 + 1267A/G were

significantly different between obese and non-obese in the control group ($p = 0.013$; $p = 0.006$).

Pociot et al. (1993) also reported that mRNA expression in insulin-dependent diabetes mellitus (IDDM) patients with homozygous GG genotype was decreased compared to the heterozygotes (AG) and homozygotes (AA).

A major strength of this study is that it was performed in a well-characterised cohort of individuals, with or without obesity; however, the main limitation of this study is the cross-sectional study design and modest sample size. In addition, subjects with Angio – had a significantly different mean age compared to the Angio + group; however, this variable was adjusted for in the logistic regression model. Also, it is possible that other lifestyle characteristics, e.g. diet, have an influence on the outcome.

In conclusion, we demonstrate the significant association of *HSP70-2* gene +1267 *HSP70-2G>A* polymorphisms with obesity and show that subjects with GG genotype or those who carried the G allele were associated with obesity, supporting further studies on evaluating the role and expression level of this emerging marker.

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Declaration of interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

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